Article

Synthesis of (2*R*,3a*R*,8a*R*)-6-Chloro-3a-hydroxy-1,2,3,3a,8,8ahexahydropyrrolo[2,3-*b*]indole-2-carboxylic Acid Methyl Ester by Reductive Cyclization

HONG, Wen-Xu(洪文旭) YAO, Zhu-Jun*(姚祝军)

State Key Laboratory of Bioorganic and Natural Products Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, Shanghai 200032, China

A synthesis of (2*R*,3*aR*,8*aR*)-6-chloro-3a-hydroxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-*b*]indole-2-carboxylic acid methyl ester (1) was achieved. An aldol reaction with Garner aldehyde, a hydroxyl introduction by Davis reagent, and a reductive intramolecular ring-closure reaction were served as the key steps. This piece of work provides a new way to synthesize the analogues of hexahydropyrrolo[2,3-*b*]indole, starting from readily available chemical substrates and inexpensive reagents.

Keywords aldol reaction, Davis oxidation, reductive ring-closure reaction, indole derivative, anticancer activity

Introduction

Pyrroloindole derivatives exist in the natural products extensively.¹⁻² They have been found to exhibit various activities including antiviral, antitoxic and antibiotic ones. Much attention was paid by synthetic chemists for these challenging structural features and biological interests. Most of existing methods^{3b-d} for generating the compounds with core structure of (2*R*, 3*aR*,8*aR*)-6-chloro-3*a*-hydroxy-1,2,3,3*a*,8,8*a*-hexahydropyrrolo[2,3-*b*] indole-2-carboxylic acid methyl ester (1) have suffered from problems with poor yields and lack of stereo-controls. One notable exception was developed by Danishefsky and coworker,^{3a} using dimethyldioxirane (DMDO) oxidation of tryptophan derivatives to give both *anti-cis* and *syn-cis* isomers of [2,3-b]pyrroloindole derivatives. More recently, a novel entrance was reported by Ley and coworkers,^{3e} using selenocyclization and oxidative deselenation of the protected tryptophan derivative to afford the *syn-cis* pyrroloindole derivatives. Herein, we report our recent elaboration on the synthesis of (2R,3aR, 8aR)-6-chloro-3a-hydroxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-b]idole-2-carboxylic acid methyl ester (1). The presented strategy and synthetic route provide the chemical com-



Figure 1 Chemical structures of (2R,3aR,8aR)-6-chloro-3a-hydroxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-*b*]indole-2-carboxylic acid methyl ester (1), chloptosin (2) and himastatin (3).

 ^{*} E-mail: yaoz@mail.sioc.ac.cn; Tel.: +8621-64163300; Fax: +8621-64166128
 Received October 16, 2003; revised November 25, 2003; accepted December 22, 2003.
 Project supported by the Major State Basic Research and Development Program of China (No. G2000077500), the National Natural Science Foundation of China (No. 20172061), the Chinese Academy of Sciences, and the Shanghai Municipal Commission of Science and Technology.

munity another choice to synthesize the *anti-syn* 3a-hydroxy-pyrroloindole derivatives. It is worthy noting here that, this kind of structures are also key subunits of some biologically important natural cyclic peptides, including chloptosin $(2)^4$ and himastatin $(3)^{3a}$ (Figure 1).

Results and discussion

The synthesis started from L-serine, which was converted into the protected amino alcohol 6 as the reference procedures.⁵ The other segment, 6-chloro-oxindole (4) could also be accessed by a direct method from 2,5-dichloronitrobenzene,⁶ in which the chlorine atom was pre-introduced in starting material to avoid the complexity in chlorination in later stage. Treatment of 4 with LDA and HMPA followed by addition of freshly prepared Garner's aldehyde 7 at −78 °C gave the aldol adducts, which were immediately subjected to MsCl and Et_3N in the same pot to afford the isomeric olefins 8 (E:Z=3.5:1).⁷ The C=C double bonds of **8a** and **8b** were then reduced by NaBH₄ in EtOH at 0 $^{\circ}$ C, providing the diastereomeric mixture 9 (ratio 1:1.6, measured by HPLC), which could not be separated by chromatography (Scheme 1). At this stage, we did not do any more endeavor to improve the diastereoselectivity because both of them will be enolated in the next step. On the other hand, we also attempted to alkylate compound 4 with fully protected 3-iodo-2-aminopropanol

Scheme 1

derivatives directly under basic conditions. Unfortunately, all these efforts failed to grant the desired intermediates.

The following hydroxyl introduction was achieved by a two-step procedure (Scheme 1). Lactam 9 was treated with LDA/HMPA at first, and then the resultant enolate was oxidized by Davis reagent $5^{8,9}$ in situ at -78 °C to give the hydroxylated lactam 10 after aqueous workup. Different Davis reagents and various bases were tested and those results are outlined in Table 1. The configurations of 10 were finally confirmed by NMR analyses of 15 in later stage (see below, Figure 2).

Selective deprotection of acetonide **10** by TsOH in MeOH afforded a mixture of **11** in 91% yield. Reaction of **11** with Boc₂O and DMAP in MeCN at room temperature afforded the compound **12a** and its diastereomer **12b**, which could be separated efficiently by regular flash column chromatography. It is worthy noting that the absolute configurations of **12a** and **12b** could not be identified at this point, and they were finally proved in the later stage by NOESY studies of compound **15** (Figure 2).

Partial reduction of lactam **12a** by LiEt₃BH in THF at -78 °C^{1a} followed by intramolecular ring-closure reaction with CBr₄-PPh₃ in ether¹⁰ at room temperature gave the desired pyrroloindole **13** in 65% yield (two steps, Scheme 2). Interestingly, of the protecting groups, which were surveyed for hydroxyl we tried, only Boc



Reaction conditions: (a) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, -78 °C, 100%. (b) (i) LDA, HMPA, **4**, THF, -78 °C; (ii) Et₃N, MsCl, CH₂Cl₂, -78 °C, 90% (20% for **8a**, and 70% for **8b**), two steps. (c) NaBH₄, EtOH, 0 °C, 73%. (d) LDA, HMPA, **5**, THF, 60%, -78 °C. (e) TsOH, MeOH, 91%. (f) Boc₂O, DMAP, MeCN, 78% (58% for **12a** and 20% for **12b**).

Ph-C	CH—N-S= 0 17	∕−сн₃	5	CI CI CI S-N-O O ⁻ S-N 0 18
Entry	Base	Davis reagent	<i>de^a</i> of 10 /%	Yield of 10 /%
1	LDA	17	15	54
2	LDA	5	24	58
3	NaHMDS	5	30	60
4	LDA/HMPA	5	49	60
5	LDA/HMPA	18	55	55

 Table 1
 The results of Davis oxidation of 9 by different bases and oxidants

^a HPLC.

was appropriate for the subsequent reduction and cyclization to the pyrroloindole **13** with LiEt₃BH. Other protecting groups (TBS, MOM) and other reduction conditions (LiAlH₄, DIBAL-H, NaBH₄, BH₃, BH₃ • SMe₂) all failed to give the desired products. Selective removal of Boc groups on the hydroxyls with 0.3 mol/L NaOH in MeOH¹¹ gave compound **14**. Oxidation of **14** with Dess-Martin periodinane followed by NaClO₂ and esterification with CH₂N₂ sequentially provided the N,N'-Boc protected methyl ester **15**. The absolute stereochemistries of **15** were finally confirmed by the NOESY experiments (Figure 2). Deprotection of both *N*-Boc groups of **15** with TFA-CH₂Cl₂ (1 : 4) afforded the amino acid methyl ester **1** in quantitative yield.

Scheme 2



Reagents and conditions: (g) (i) LiBEt₃H, THF; (ii) CBr₄, PPh₃, ether, 65% in two steps. (h) NaOH, MeOH, 93%. (i) Dess-Martin periodinane CH₂Cl₂; (ii) NaH₂PO₄, 2-methyl-2-butene, NaClO₂, *t*-butanol, H₂O; (iii) CH₂N₂, Et₂O, 75% in three steps. (j) TFA, CH₂Cl₂, 100%.



Figure 2 Chemical shifts and NOESY of compound 15.

Conclusion

In summary, a stereoselective synthesis of (2R,3aR, 8aR)-6-chloro-3a-hydroxy-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-*b*]indole-2-carboxylic acid methyl ester (1) was described. An aldol reaction with Garner aldehyde, a hydroxyl introduction by Davis reagent, and a reductive intramolecular ring-closure reaction were successfully served as the key steps. It provides a new route to obtain these important pyrroloindole derivatives by chemical synthesis, starting from readily available chemical substrates and inexpensive reagents.

Experimental

All reactions were carried out under argon or nitrogen in oven-dried glassware using standard gastight syringes, cannulas, and septa. Solvents and reagents were purified and dried by standard methods prior to use. Optical rotations were measured at room temperature. IR spectra were recorded on an FT-IR instrument. ¹H NMR spectra were recorded at 300 MHz and ¹³C NMR spectra were recorded at 75 MHz. Flash column chromatographies were performed on silica gel (10—40 μ m) using mixtures of petroleum ether and acetate as eluents.

Compounds 8a and 8b To a solution of oxalyl chloride (2.83 mL, 32.4 mmol) in dry CH_2Cl_2 (40 mL) at -75 °C was added DMSO (3.7 mL, 52 mmol) in dry CH_2Cl_2 (5 mL) in 15 min. After being stirred at -75 °C for an additional 15 min, alcohol **6** (5.0 g, 21.6 mmol) in dry CH_2Cl_2 (40 mL) was added over 5 min. After the reaction was stirred at -75 °C for 2 h, dry Et₃N (8.2 mL, 58.4 mmol) was added and the mixture was slowly warmed to room temperature. Saturated NH₄Cl was added and the mixture was extracted with Et₂O (50 mL). The combined Et₂O layers were washed with brine, dried over Na₂SO₄ and concentrated to yield a yellow oil **7**, which was used directly for the next step.

To a solution of diisopropylamine (4.2 mL, 30 mmol) in dry THF (100 mL) at -75 °C was added dropwise *n*-BuLi (2.5 mol/L in hexane, 10 mL, 25 mmol). After 30 min, HMPA (3.5 mL, 20 mmol) was added dropwise and the mixture was stirred for an additional 30 min. Indole derivative **4** (1.67 g, 10 mmol) in dry THF (40 mL) was added over 5 min. After 30 min, the above aldehyde **7** in dry THF (40 mL) was added dropwise and the reaction was stirred for 1.5 h. Saturated NH₄Cl was added at -78 °C to quench the reaction. The mixture was warmed to room temperature and extracted with EtOAc (50 mL×3). The combined organic layers were washed with water (50 mL×2) and brine (50 mL), dried over Na₂SO₄ and concentrated to give a red oil, which was used directly for the next step.

To a solution of the above intermediate in CH₂Cl₂ (30 mL) at -78 °C was added Et₃N (4.1 mL, 30 mmol) and MsCl (1.2 mL, 15 mmol), and the solution was stirred for 2 h. Saturated NH₄Cl was added at -78 °C and the mixture was warmed to r.t. and extracted with CH_2Cl_2 (50 mL×3). The combined organic layers were washed with water (50 mL \times 3) and brine (50 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography to give 8a and 8b as yellow solids (760 mg of 8a and 2.64 g of 8b, in 90% yield totally). Data for 8a: m.p. 208-210 °C; $[\alpha]_{D}^{25}$ =51.9 (*c* 0.856, CH₃Cl); ¹H NMR (300 MHz, CDCl₃) δ: 8.96 (brs, 1H), 7.34–6.72 (m, 4H), 5.86 (brs, 1H), 4.39 (brs, 1H), 3.84 (d, J=8.2 Hz, 1H), 1.73-1.33 (m, 15H); IR (KBr) v: 3240, 2976, 1708 cm^{-1} ; MS (EI) *m/z*: 322 (M⁺+1-57). Anal. calcd for C₁₉H₂₃ClN₂O₄: C 60.23, H 6.12, N 7.39; found C 60.12, H 6.29, N 6.97. Data for **8b**: m.p. 152–153 °C; $[\alpha]_{D}^{25}$ +9.8 (c 1.377, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ : 8.89 (brs, 1H), 7.36 (d, J=8.1 Hz, 1H), 7.01-6.83 (m, 3H), 5.17 (brs, 1H), 4.30 (dd, J=9.3, 6.9 Hz, 1H), 3.88 (dd, J=9.0, 3.0 Hz, 1H), 1.70-1.28 (m, 15H); IR (KBr) v: 2982, 1716, 1613, 1392 cm⁻¹; MS (EI) *m/z*: 379 (M⁺ +1). Anal. calcd for $C_{19}H_{23}ClN_2O_4$: C 60.23, H 6.12, N 7.39; found C 60.63, H 6.38, N 6.99.

Compound 9 To a solution of NaBH₄ (860 mg, 22.6 mmol) in absolute EtOH (150 mL) at 0 $^{\circ}C$ was added a solution of 8a and 8b (4.3 g, 11.3 mmol) in absolute EtOH (10 mL) and the mixture was stirred for 2 h at 0 °C. The reaction was quenched by addition of saturated NaHCO₃ at 0 °C and EtOH was evaporated in *vacuo*. The mixture was extracted by EtOAc (50 mL \times 3) and the combined EtOAc layers were washed with brine (50 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography to give **9** as a white solid (3.14 g, 73%, 23% *de* by HPLC). ¹H NMR (300 MHz, CDCl₃) δ : 9.64 (brs, 1H), 7.37-6.99 (m, 3H), 4.38-4.35 (m, 1H), 4.05-3.89 (m, 2H), 3.50-3.40 (m, 1H), 2.16-2.06 (m, 2H), 1.55 -1.45 (m, 15H); MS (EI) m/z: 380 (M⁺); HRMS (EI) calcd for $C_{19}H_{25}ClN_2O_4$ [M⁺] 380.1503, found 380.1455.

Compound 10 To a solution of diisopropylamine (16 mg, 0.158 mmol) in dry THF (1 mL) was added *n*-BuLi (1.6 mol/L in hexane, 100 μ L, 0.16 mmol) at -78 °C. After 30 min, HMPA (29 mg, 0.16 mmol) was added and the mixture was stirred for an additional 30 min. Compound **9** (50 mg, 0.13 mmol) in dry THF (1 mL) was added dropwise. After the reaction was stirred for 30 min, Davis reagent **5** (60 mg, 0.26 mmol) in dried THF (1 mL) was added. The reaction was continued to stir for 2 h at -78 °C. The reaction was quenched by addition of saturated NH₄Cl and then warmed to r.t. The mixture was extracted with EtOAc (50 mL×3). The combined organic layers were washed with brine (50 mL), dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The residue was purified by flash column chromatography to give **10** as a white solid (32 mg, 60%, 49% *de* by HPLC). ¹H NMR (300 MHz, CDCl₃) δ : 8.75 (brs, 1H), 7.33—6.77 (m, 3H), 4.58—4.37 (m, 1H), 4.09—3.68 (m, 2H), 2.43—1.93 (m, 2H), 1.54—1.34 (m, 15H) (one of the active protons was not observed in the spectrum); MS (EI) *m/z*: 381 (M⁺-15); HRMS (EI) calcd for C₁₈H₂₃ClN₂O₅ [M⁺+1–15] 382.1260, found 382.1310.

Compound 11 To a solution of diastereomeric mixture **10** (453 mg, 1.14 mmol) in MeOH (10 mL) was added TsOH hydrate (12 mg, 0.063 mmol), and the mixture was stirred at r.t. overnight. The solution was concentrated *in vacuo* and the residue was purified by flash column chromatography to give **11** as a white solid (413 mg, 91%). ¹H NMR (300 MHz, DMSO- d_6) δ : 10.35 (brs, 1H), 7.25—7.18 (m, 1H), 6.98—6.96 (m, 1H), 6.74—6.70 (m, 1H), 5.92—5.89 (m, 1H), 4.63—4.61 (m, 1H), 3.25 (brs, 1H), 3.13 (brs, 2H), 2.17—2.10 (m, 2H), 1.33—1.19 (m, 9H) (one of the active protons was not observed in the spectrum); MS (ESI) *m/z*: 357 (M⁺+1). Anal. calcd for C₁₆H₂₁ClN₂O₅: C 53.86, H 5.93, N 7.85; found C 53.56, H 6.00, N 7.49.

To a solution of compound 11 Compound 12 (1.0 g, 2.8 mmol) in MeCN (10 mL) was added DMAP (40 mg, 0.3 mmol) and Boc₂O (1.83 g, 8.4 mmol) at r.t. The reaction was stirred overnight and quenched with sat. NH₄Cl. The mixture was diluted with EtOAc (30 mL) and the layers were separated. The aqueous layer was extracted with EtOAc (30 mL \times 3) and the combined organic layers were washed with water (50 mL) and brine (50 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. Purification on silica gel provided 12 as a white solid (1.64 g of 12a, 0.41 g of 12b, 78% yield totally). Data for **12a**: m.p. 143—145 °C; ¹H NMR (300 MHz, CDCl₃) δ : 7.96 (s, 1H), 7.40 (d, J =8.1 Hz, 1H), 7.19 (dd, J=8.1, 1.2 Hz, 1H), 4.57 (d, J=8.4 Hz, 1H), 4.04 (d, J=8.1 Hz, 2H), 3.94-3.90 (m, 1H), 2.32-2.28 (m, 1H), 2.07 (d, J=11.4 Hz, 1H), 1.65 (s, 9H), 1.46 (s, 9H), 1.39 (s, 9H), 1.34 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ : 172.0, 154.5, 153.0, 150.5, 148.4, 140.6, 135.8, 124.8, 124.7, 123.8, 116.4, 85.0, 84.0, 82.5, 69.2, 45.0, 38.5, 28.2, 27.9, 27.6, 27.4; IR (KBr) v: 3381, 2981, 1770, 1745, 1613, 1526, 1283 cm^{-1} ; MS (ESI) m/z: 674 (M + NH₄⁺); HRMS (ESI) calcd for $C_{31}H_{45}ClN_2NaO_{11}$ [M⁺+Na] 679.2604, found 679.2611.

Compound 13 To a solution of **12a** (36 mg, 0.055 mmol) in dry THF (3 mL) at -78 °C was added LiBEt₃H (1 mol/L in THF, 0.12 mL, 0.12 mmol). The reaction mixture was stirred at -78 °C for 30 min before it was quenched with sat. NaHCO₃ and diluted with EtOAc (5 mL). The layers were separated and the aqueous layer was extracted with EtOAc (5 mL×2). The combined organic layers were washed with water (5 mL) and brine (5 mL), dried over Na₂SO₄, filtered and

concentrated *in vacuo*. The residue was directly used in the next step.

To a solution of the above residue in Et_2O (5 mL) was added CBr₄ (40 mg, 0.12 mmol) and PPh₃ (31 mg, 0.12 mmol). The reaction mixture was stirred at r.t. for 30 min before it was diluted with EtOAc (5 mL) and quenched with sat. NaHCO₃. The layers were separated and aqueous layer was extracted with EtOAc (5 mL \times 3). The combined organic layers were washed with water (5 mL) and brine (5 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. Purification on silica gel provided 13 as a waxy solid (19 mg, 65% in two steps). $[\alpha]_{D}^{25}$ -44.5 (c 0.867, CH₃Cl); ¹H NMR (300 MHz, CDCl₃) δ :7.75 (s, 1H), 7.30 (d, J=8.4 Hz, 1H), 7.06 (dd, J=7.8, 1.8 Hz, 1H), 6.08 (s, 1H), 4.36-4.33 (m, 1H), 3.89 (dd, J=10.2, 5.1 Hz, 1H), 3.13 (t, J=9.9 Hz,1H), 2.56 (dd, J=13.8, 9.0 Hz, 1H), 2.43 (dd, J=13.5, 1.5 Hz, 1H), 1.59 (s, 9H), 1.49 (s, 9H) 1.40 (s, 9H) (one active proton was not observed in the spectrum); ¹³C NMR (75 MHz, CDCl₃) δ : 154.0, 152.9, 152.2, 142.9, 136.2, 131.9, 124.1, 123.8, 117.6, 84.9, 83.5, 82.5, 82.2, 81.1, 66.7, 57.2, 39.7, 28.5, 28.3, 28.2, 27.7; IR (film) v: 3416, 2925, 1719, 1598 cm⁻¹; MS (ESI) m/z: 541 (M⁺+1); HRMS (ESI) calcd for C₂₆H₃₇O₈- N_2 ClNa [M⁺+Na] 563.2131, found 563.2137.

Compound 14 To a solution of 13 (42 mg, 0.078 mmol) in MeOH (10 mL) was added NaOH solution (0.6 mol/L in MeOH, 10 mL, 6.0 mmol). The reaction mixture was stirred for 24 h at r.t., and then diluted with EtOAc and neutralized with aqueous HCl (1 mol/L). MeOH was evaporated under reduced pressure and the residue extracted with EtOAc (5 mL \times 3). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. Chromatography on silica gel provided 14 (32 mg, 93%) as a waxy solid. $[\alpha]_{D}^{25}$ -72.7 (c 0.787, CH₃Cl); ¹H NMR (300 MHz, CDCl₃) δ : 7.65 (s, 1H), 7.30 (d, J= 7.8 Hz, 1H), 7.07 (dd, J=8.4, 1.8 Hz, 1H), 6.10 (s, 1H), 4.30 (d, J=6.3 Hz, 1H), 3.07-3.05 (m, 2H), 2.62-2.51 (m, 1H), 2.28 (dd, J=13.2, 1.8 Hz, 1H), 1.57 (s, 9H), 1.48 (s, 9H) (two active protons were not observed in the spectrum); ¹³C NMR (75 MHz, CDCl₃) δ : 155.7, 152.2, 142.6, 135.9, 132.3, 124.0, 117.9, 84.6, 83.5, 82.6, 81.9, 65.6, 60.8, 39.6, 28.4, 28.2; IR (film) v: 3399, 2982, 2927, 2847, 1723, 1602, 1477, 1395 cm MS (EI) m/z: 440 (M⁺); HRMS (MALDI) calcd for $C_{21}H_{29}CIN_2NaO_6 [M^++Na] 463.1606$, found 463.1628.

Compound 15 To a solution of **14** (20 mg, 0.046 mmol) in $CH_2Cl_2(2 \text{ mL})$ was added Dess-Martin periodinane (28 mg, 0.66 mmol) at r.t. The reaction mixture was stirred for 30 min and quenched with sat. NaHCO₃ containing Na₂S₂O₃. After being stirred for 20 min, the layers were separated. The aqueous layer was extracted with EtOAc (5 mL×3) and the combined extracts were washed with sat. NaHCO₃ (5 mL), dried over Na₂SO₄ and evaporated *in vacuo*. The residue was used directly in the next step.

To a solution of the above aldehyde in *t*-butanol and water (4:1, V/V, 10 mL) at 0 °C was successively

added 2-methyl-2-butene (13 mg, 0.184 mmol), NaH₂PO₄ (6 mg, 0.048 mmol) and NaClO₂ (12.5 mg, 0.138 mmol). After being stirred at room temperature for 1 h, the reaction mixture was quenched with NH₄Cl (sat.) and extracted with EtOAc (5 mL \times 3). The organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo* to give the acid as a white solid, which was used directly in the next step.

To a solution of the above acid in Et₂O (15 mL) at 0 °C, CH₂N₂ in Et₂O was added until the solution turned to yellow. After being stirred for an additional 20 min, excess CH₂N₂ was destroyed by adding glacial acetic acid. The mixture was concentrated in vacuo and the residue was purified by flash column chromatography to give 15 as a white solid (15 mg, 75% in three steps). m.p. 162—164 °C; $[\alpha]_{D}^{25}$ -33.3 (c 0.44, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ : 7.27 (s, 1H), 7.19 (d, J =8.1 Hz, 1H), 7.01 (dd, J=8.1, 1.8 Hz, 1H), 6.03 (s, 1H), 4.62 (t, J=5.3 Hz, 1H), 3.24 (s, 3H), 2.75 (s, 1H), 2.71 $(d, J=5.7 \text{ Hz}, 2\text{H}), 1.57 (s, 9\text{H}), 1.44 (s, 9\text{H}); {}^{13}\text{C NMR}$ (75 MHz, CDCl₃) δ: 171.4, 152.2, 144.4, 136.4, 130.4, 124.4, 123.4, 117.6, 84.3, 82.5, 82.1, 82.1, 81.1, 59.4, 52.0, 39.7, 28.3, 28.2; IR (KBr) v: 3445, 1729, 1679, 1601, 1478, 1401, 1365, 1239, 1169, 809, 747 cm⁻¹; MS (ESI) m/z: 491 (M⁺+Na); HRMS (ESI) calcd for $C_{22}H_{29}ClN_2NaO_7$ [M⁺+Na] 491.1556, found 491.1541.

To a solution of **15** (9 mg, 0.019 Compound 1 mmol) in CH₂Cl₂ (4 mL) was added TFA (1 mL) at 0 °C. After being stirred at r.t. for 5 h, the reaction mixture was diluted with CH₂Cl₂ (5 mL) and evaporated in vacuo. The residue was dissolved in CH₂Cl₂ (5 mL), neutralized to pH 7-8 by NaHCO₃ (sat.) and extracted with CH_2Cl_2 (5 mL×3). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography to give 1 as a white solid in quantitative yield. m.p. 163–165 °C; $[\alpha]_{D}^{25}$ –57.4 (c 0.14, CHCl₃). ¹H NMR (300 MHz, DMSO- d_6) δ : 7.04 (d, J=7.5 Hz, 1H), 6.52 (dd, J=8.1, 2.4 Hz, 1H), 6.39 (d, J=1.5 Hz, 1H), 6.32 (s, 1H), 5.63 (s, 1H), 4.71 (d, J =1.8 Hz, 1H), 3.95 (t, J=7.2 Hz, 1H), 3.42 (s, 3H), 2.30 (dd, J=12.9, 7.2 Hz, 1H), 2.07 (dd, J=12.6, 6.9 Hz, 1H) 1.33 (s, 1H); IR (film) v: 3270, 2955, 1735, 1486, 1442, 1311, 1230, 1138, 1106, 805 cm⁻¹; MS (ESI) m/z: 291 [M⁺+Na]; HRMS (ESI) calcd for C₁₂H₁₃ClN₂NaO₃ [M⁺+Na] 291.0512, found 291.0507.

References

 (a) Depew, K. M.; Marsden, S. P.; Zatorska, D.; Zatorski, A.; Bornmann, W. G.; Danishefsky, S. J. J. Am. Chem. Soc. 1999, 121, 11953.

(b) Overman, L. E.; Paone, D. V. J. Am. Chem. Soc. 2001, 123, 9465.

(c) Overman, L. E.; Govek, S. P. J. Am. Chem. Soc. 2001, 123, 9468.

- 2 Daly, J. W. J. Med. Chem. 2003, 46, 445.
- 3 (a) Kamenecka, T. M.; Danishefsky, S. J. Chem.-Eur. J. 2001, 7, 41.

(b) Fukui, Y.; Somei, M. Heterocycles 2001, 55, 2055.

(c) Anthoni, U.; Christophersen, C.; Nielsen, P. H.; Christoffersen, M. W.; Sorensen, D. *Acta Chem. Scand.* **1998**, *52*, 958.

(d) Sakai, A.; Tani, H.; Aoyama, T.; Shioiri, T. *Synlett* **1998**, 257.

(e) Ley, S. V.; Cleator, E.; Hewitt, P. R. *Org. Biomol. Chem.* **2003**, (1), 3492.

- 4 Umezawa, K.; Ikeda, Y.; Uchihata, Y.; Naganawa, H.; Kondo, S. J. Org. Chem. **2000**, 65, 459.
- 5 Avenoza, A.; Cativiela, C.; Corzana, F; Peregrina, J. M.; Zurbano, M. M. *Synthesis* **1997**, 1146.

- 7 Ma, D.; Wu, Q. Tetrahedron Lett. 2000, 41, 9089.
- 8 Davis, F. A.; Haque, M. S.; Ulatowski, T. G.; Towson, J. C. J. Org. Chem. 1986, 51, 2402.
- 9 Kato, T.; Tomita, N.; Hoshikawa, M.; Ehara, K.; Shima, J.; Takahashi, N.; Sugiyama, H. *Heterocycles* **1998**, *47*, 497.
- Marino, J. P.; Bogdan, S.; Kimura, K. J. Am. Chem. Soc. 1992, 114, 5566.
- 11 Namba, K.; Shinada, T.; Tetramoto, T.; Ohfune, Y. J. Am. Chem. Soc. **2000**, *122*, 10708.

(E0310162 PAN, B. F.)